



**(For scientific research use only, not for clinical diagnosis!)**

## **Fish Platelet-Derived Growth Factor (PDGF)**

**ELISA Kit Instructions for Use Product No.:**

**BY-EF990845 Specifications: 48T/96T**

**Detection range: 250 pg/mL– 8000 pg/mL.**

**Sensitivity:** The lowest detectable dose is less than 10 pg/mL.

**Precision:** intra-batch variation coefficient CV% is less than 10%; inter-batch variation coefficient CV% is less than 15%.

**Recovery rate:** The recovery rate is between 85%-115%.

**Specificity:** This kit recognizes natural and recombinant fish platelet-derived growth factor (PDGF) and has no crossover with structural analogs. **Stability:** Stored at 2℃-8℃, validity period is 6 months.

**Purpose:** Used to detect the concentration of fish platelet-derived growth factor (PDGF) in samples such as serum, plasma, cell culture supernatant and tissue.

**Please read the instructions carefully before use. If you have any questions, please contact us through the following methods: Official hotline: 025-5229-8998 Sales department phone: 13914481711 Technical phone: 15950492658**

**Company website: [www.byabscience.cn](http://www.byabscience.cn) For the specific shelf life, please refer to the outer packaging label of the kit. Please use the kit within the shelf life.**

When contacting us, please provide the product number and production date (see box label) so that we can serve you more efficiently.

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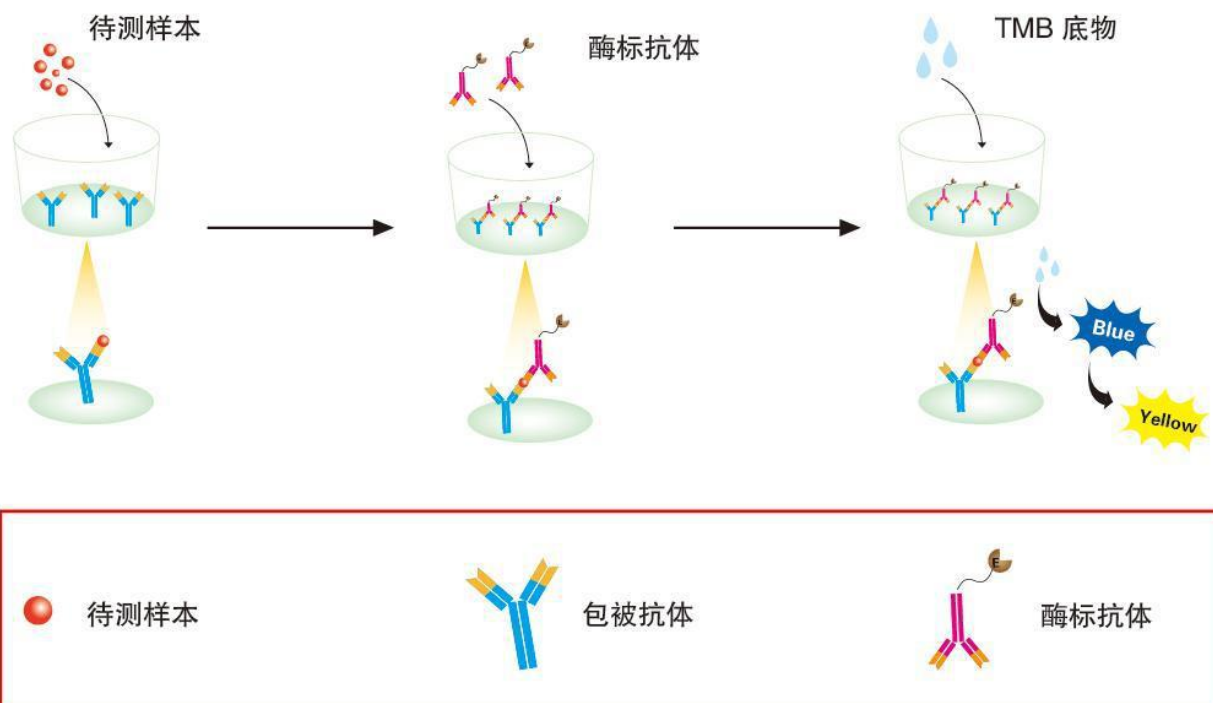
**Supervision phone number:**



## Experimental principle

This kit uses a double-antibody sandwich enzyme-linked immunosorbent assay (ELISA). In the microwell enzyme plate pre-coated with anti-fish platelet-derived growth factor (PDGF) antibody (solid-phase antibody), add the fish platelet-derived growth factor (PDGF) calibrator and the sample to be tested, and then add HRP-labeled Anti-fish platelet-derived growth factor (PDGF) antibody (enzyme-labeled antibody), after incubation and sufficient washing, removes unbound components, and forms a sandwich of solid-phase antibody-antigen-enzyme-labeled antibody on the solid surface of the microplate complex. Add substrates A and B. The substrates are catalyzed by HRP to produce a blue product, which is finally converted to yellow under the action of the stop solution (acidic solution). The absorbance (OD value) is measured at the 450nm wavelength of a microplate reader. The absorbance (OD value) is positively correlated with the concentration of fish blood platelet-derived growth factor (PDGF) in the sample to be tested. By fitting the calibrator curve, the concentration of fish platelet-derived growth factor (PDGF) in the sample can be calculated.

## Experimental schematic diagram



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**Kit components and storage: Store unopened kits at 2-8 degrees Celsius. Do not use expired kits.**

| Components              | 48-well configuration | 96-well configuration | Store after opening  |
|-------------------------|-----------------------|-----------------------|----------------------|
| Pre-coated enzyme plate | 48T                   | 96T                   | <b>2-8°C 14 days</b> |
| Standard product        | 0.3mL*6 tubes         | 0.3mL*6 tubes         | <b>2-8°C 14 days</b> |
| sample diluent          | 3ml                   | 6ml                   | 2-8°C 180 days       |
| HRP labeled antibodies  | 5ml                   | 10ml                  | <b>2-8°C 14 days</b> |
| Chromogenic substrate A | 3ml                   | 6ml                   | 2-8°C 180 days       |
| Chromogenic substrate B | 3ml                   | 6ml                   | 2-8°C 180 days       |
| stop solution           | 3ml                   | 6ml                   | 2-8°C 180 days       |
| <b>20×Lotion</b>        | 15ml                  | 25ml                  | 2-8°C 180 days       |
| sealing film            | 2 sheets              | 2 sheets              |                      |
| manual                  | 1 serving             | 1 serving             |                      |
| Ziplock bag             | 1                     | 1                     |                      |

The concentrations of calibrators are: 8000, 4000, 2000, 1000, 500 and 250 pg/mL.

Note: 1: Before use, please check whether the label and quantity of

the reagents in the kit are consistent with the table.

2: If the components of the kit need to be used again, please ensure that they have

not been contaminated since the last use. 3: If the enzyme plate is not used up in a

single time, remember to seal it and store it at 2-8°C.

**Prepare your own test equipment required for the test (not provided, but can assist in**

1) Microplate reader capable of detecting

absorbance at 450 nm 2) Pipette, pipette tip,

and sample addition tank 3) 37°C incubator

or water bath 4) Test tubes, centrifuge tubes,

measuring cylinders, etc. for preparing

reagents 5) Distilled water or deionized

water Ionized water

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6) Vortex shaker, microplate shaker

**Notes 1) For scientific research use only,  
not for clinical diagnosis.**

- 2) Use within the validity period marked on the kit. Expired products must not be used.
- 3) Do not mix with kits or components from other manufacturers. Use the sample diluent provided with the kit.
- 4) If the sample value is higher than the highest standard concentration value, please dilute the sample appropriately and then re-measure.
- 5) Human anti-mouse and other heterophilic antibodies present in the sample to be tested will interfere with the test results. Please eliminate this factor before testing.
- 6) The test results obtained by other methods are not directly comparable to the test results of this kit.
- 7) Please wear a lab coat and latex gloves for protection during the test. Especially when testing blood or other body fluid samples, please follow the national biological laboratory safety protection regulations.
- 8) Carry out incubation strictly according to the specified time and temperature to ensure accurate results. All reagents must reach room temperature 20-25°C before use. Store reagents refrigerated immediately after use.
- 9) Improper plate washing can lead to inaccurate results. Make sure to absorb as much liquid as possible from the wells before adding substrate. Do not allow the microwells to dry out during incubation.
- 10) Eliminate residual liquid and fingerprints on the bottom of the plate, otherwise it will affect the OD value.
- 11) The substrate chromogenic solution should be colorless or very light in color.
- 12) Avoid cross-contamination of reagents and specimens to avoid erroneous results.

13) Avoid direct exposure to strong light during storage and incubation.

14) The microplate reader used for detection needs to be equipped with a filter capable of detecting a wavelength of  $450\pm 10\text{nm}$ , and the optical density range is between 0-3.5. It is recommended to preheat 15 minutes in advance before use.

15) The EP tubes and tips used in the test are single-use and are strictly prohibited from mixing.

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## Sample preparation and storage

The following lists only general guidelines for sample collection and preservation. During the collection and storage of all samples, sodium azide shall not be used as a preservative. If the sample is not analyzed immediately, it should be aliquoted and stored frozen, and repeated freezing and thawing should be avoided.

**Cell culture supernatant - centrifuge to remove precipitate, analyze immediately or aliquot and store frozen at -20°C.**

**Serum - Collect blood in a clean test tube, coagulate at room temperature for 30 minutes, centrifuge at 2000×g for 20 minutes, and collect serum. Analyze immediately or aliquot and store frozen at -20°C.**

**Plasma—anticoagulate with heparin, citrate, or EDTA, and centrifuge at 2000×g for 20 minutes at 2-8°C within 30 minutes of blood draw. To eliminate the influence of platelets, it is recommended to further centrifuge at 10,000 × g for 10 minutes at 2-8°C. Analyze immediately or aliquot and store frozen at -20°C.**

**Cell lysis buffer - For adherent cells, remove the culture medium and wash with PBS, normal saline or serum-free culture medium. Add an appropriate amount of lysis solution and pipet several times with a gun to fully contact the lysate and cells. Typically after 10 seconds, cells are lysed. For suspended cells, collect the cells by centrifugation and wash them with PBS, physiological saline or serum-free culture medium. Add an appropriate amount of lysis solution, blow the cells with a gun, and flick them with your fingers to fully lyse the cells. After full lysis, centrifuge at 10000-14000×g for 3-5 minutes and take the supernatant. Analyze immediately or aliquot and store frozen at -20°C.**

**Tissue homogenate - rinse the tissue with pre-cooled PBS (0.01M, pH=7.4) to remove residual blood (lysed red blood cells in the homogenate will affect the measurement results), weigh and cut the tissue into pieces. Mix the minced tissue with the corresponding volume of PBS (generally**

at a weight-to-volume ratio of 1:9, for example, 1g of tissue sample corresponds to 9mL of PBS. The specific volume can be adjusted appropriately according to experimental needs and recorded. It is recommended to add Protease inhibitor) was added to a glass homogenizer and ground thoroughly on ice. In order to further lyse tissue cells, the homogenate can be sonicated or frozen and thawed repeatedly. Finally, centrifuge the homogenate at  $5000 \times g$  for 5 to 10 minutes, and take the supernatant for detection.

**Urine - Collect in sterile tubes and centrifuge at  $2000 \times g$  for 20 minutes. Carefully collect the supernatant. If a precipitate forms, centrifuge again.**

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**Reagent preparation 1. Before use, all components must be rewarmed**

**for at least 60 minutes to ensure sufficient rewarming to room temperature.**

2. Concentrated washing liquid: The concentrated washing liquid taken out from the refrigerator will produce crystals. This is a normal phenomenon. Heating in a water bath will completely dissolve the crystals. Concentrated detergent and distilled water, dilute 1:20, that is, 1 part of concentrated detergent, add 19 parts of distilled water. 3. Substrate: Substrate solutions A and B, mix thoroughly at a volume of 1:1 before use, and use within 15 minutes after mixing.

**Operating procedures: Return all reagents and components to room temperature first. For standards, quality control materials and samples, it is recommended to make duplicate holes.**

1. Prepare the working solution of various components of the kit according to the method described in the previous instructions.
2. Take out the required slats from the aluminum foil bag, seal the remaining slats in a ziplock bag and return it to the refrigerator.
3. Set up standard wells, 0 value wells, blank wells and sample wells. Add 50  $\mu\text{L}$  of standards of different concentrations to each of the standard wells. Add 50  $\mu\text{L}$  of sample diluent to the 0 value well. Do not add any to the blank well. Add 50  $\mu\text{L}$  of the sample to be tested to the sample well. .
4. In addition to the blank wells, add 100  $\mu\text{L}$  of horseradish peroxidase (HRP)-labeled detection antibody to the standard wells, 0 value wells and sample wells.
5. Cover the reaction plate with sealing film and incubate in a 37°C water bath or incubator in the dark for 60 minutes.
6. Uncover the sealing film, discard the liquid, pat dry on absorbent paper, fill each well with washing solution, let stand for 20 seconds, shake off the washing solution, pat dry on absorbent paper, repeat this

5 times. If you use an automatic plate washer, please wash the plate according to the plate washer operating procedure. Adding a soaking program for 30 seconds can improve the detection accuracy. After washing the plate and before adding substrate, pat the reaction plate dry on clean, lint-free paper. (Tip: In order to obtain ideal experimental results, the residual liquid must be completely removed. After washing the plate, please proceed to the next step immediately and do not let the microplate dry.) 7. Mix substrates A and B at a volume of 1:1 Mix thoroughly and add 100  $\mu$ L of substrate mixture to all wells. Cover the reaction plate with sealing film and incubate in a 37°C water bath or incubator in the dark for 15 minutes.

8. Add 50  $\mu$ L of stop solution to all wells, and read the absorbance (OD value) of each well on a 450nm wavelength microplate reader.

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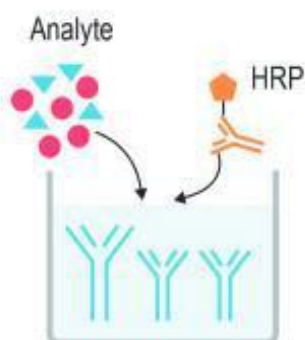
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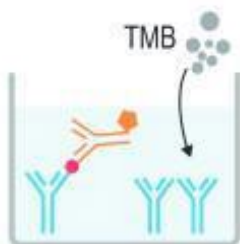
**[Operation flow chart]**



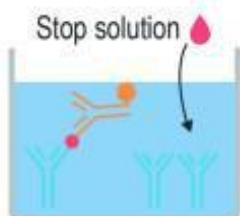
1. 对应板孔中加入50 $\mu$ L标准品工作液或样本后，立即每孔加入100 $\mu$ L HRP酶标抗体工作液，37 $^{\circ}$ C孵育60分钟



2. 弃掉板内液体，洗板5次



3. 每孔加入底物A溶液50 $\mu$ L，底物B溶液50 $\mu$ L



4. 每孔加入50 $\mu$ L终止液

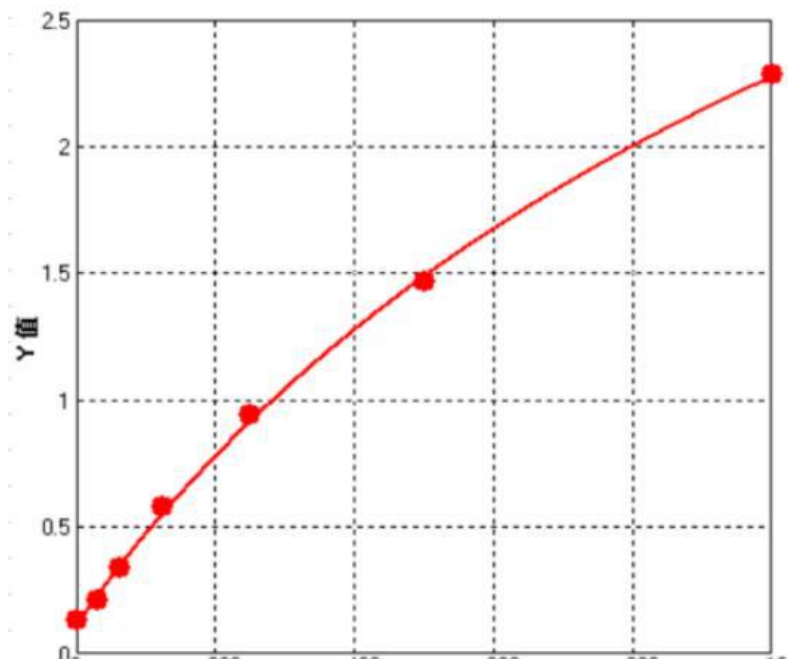


5. 立即在450nm波长下读数，处理数据



## Result calculation

9. Use the concentration of the standard substance as the abscissa and the corresponding absorbance (OD value) as the ordinate. Use computer software and four-parameter Logistic curve fitting (4-pl) to create a standard curve equation. Through the absorbance (OD value) of the sample value), use the equation to calculate the concentration value of the sample. [Calculate using ELISA Calc software. It is recommended to use four-parameter fitting for the standard curve, but it is not the only fitting method]
10. If the sample is diluted, the concentration value measured by the above method must be multiplied by the dilution factor to determine the final value of the sample. concentration. Note: Experimenters need to establish a standard curve based on their own experiments. For each test, a standard curve must be established for each enzyme plate. The following curves are for reference only!



(Schematic diagram of the music, for reference only)

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**[Problem Analysis]** If the experimental results are not good, please take pictures of the color development results in time, save the experimental data, keep the used laths and unused reagents, and then contact our company's technical support to solve the problem for you. At the same time, you can also refer to the following information:

**[Questions and Answers]**

| Problem description                | Possible reasons                                | Corresponding countermeasures                                                                                                    |
|------------------------------------|-------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------|
| standard curve gradient difference | Incorrect liquid aspiration or                  | Check pipettes and tips                                                                                                          |
|                                    | Equilibration time is too short                 | Ensure sufficient balancing time                                                                                                 |
|                                    | Incomplete washing                              | Ensure the washing time and number of washes and the amount of liquid added to each hole                                         |
| Very weak or colorless             | Incubation time too short                       | Ensure adequate incubation time                                                                                                  |
|                                    | The experimental temperature is incorrect       | Use recommended experimental temperatures                                                                                        |
|                                    | Insufficient reagent volume or missing addition | Check the liquid aspirating and adding process to ensure that all reagents are added in order and in                             |
|                                    | Incorrect dilution                              |                                                                                                                                  |
|                                    | Enzyme label inactivation or substrate failure  | Mix enzyme conjugate and substrate and check by rapid color development                                                          |
| Reading value is low               | Microplate reader settings are incorrect        | Check the wavelength and filter                                                                                                  |
|                                    |                                                 | Turn on the microplate reader and preheat it in advance                                                                          |
| Large coefficient of variation     | Adding fluid incorrectly                        | Check the filling situation                                                                                                      |
| High background value              | The working concentration of the                | Use the recommended dilution                                                                                                     |
|                                    | Incomplete washing of enzyme plate              | Ensure that each step of cleaning is complete; if using an automatic plate washer, please check whether all outlets are blocked; |
|                                    | The lotion is contaminated                      | Prepare fresh lotion                                                                                                             |
| Low sensitivity                    | Improper storage of ELISA kits                  | Store relevant reagents according to                                                                                             |
|                                    | Not terminated before reading                   | Stop solution should be added to                                                                                                 |



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## statement

1. Due to the current conditions and scientific and technological level, it is not possible to conduct comprehensive identification and analysis of all raw materials.  
This product may have certain quality and technical risks.
2. This kit removes/reduces some endogenous interfering factors in biological samples during the development process. Not all possible influencing factors have been removed.
3. The final experimental results are closely related to factors such as the effectiveness of the reagents, the relevant operations of the experimenter, and the experimental environment at the time. Our company is only responsible for the kit itself and is not responsible for the sample consumption caused by the use of the kit.  
Please use The user should fully consider the possible usage of the sample and reserve sufficient samples before use.
4. In order to achieve good experimental results, please only use the reagents provided in our company's kits, do not mix products from other manufacturers, and operate in strict accordance with the instructions.
5. Due to incorrect reagent preparation and microplate reader parameter settings during the operation, abnormal results may occur. Please read the instructions carefully and adjust the instrument before the experiment.
6. Even if operated by the same personnel, different results may be obtained in two independent experiments. In order to ensure the reproducibility of the results, it is necessary to control every step of the experimental process.

7. The kits will undergo strict quality inspection before shipment. However, due to factors such as transportation conditions, differences in experimental equipment, etc., user test results may be inconsistent with factory data.

8. This kit has not been compared with similar kits from other manufacturers or products using different methods to detect the same target, so inconsistent test results cannot be ruled out.

9. The kit is for research use only. If it is used for clinical diagnosis or any other purpose, our company will not be responsible for any problems arising therefrom, nor will we assume any legal liability.

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